

Computational Biology (BIOSC 1540)

Lecture 18: Ligand-based drug design

Nov 7, 2024

Announcements

- A07 is due Thursday by 11:59 pm
- CSB exam is next Thursday (Nov 14)
 - Study guide will be posted tonight or tomorrow
 - We will have a review session on Tuesday (Nov 12)
 - Request DRS accommodations if needed
- Project will be due Dec 10
- OMETs will be coming out soon
- Attending our optional Python lectures are strongly recommended if you are taking simulation on modeling

After today, you should have a better understanding of

The basic principles of ligand-based drug design and how it differs from structure-based approaches.

Structural insight into a disease is a privilege

Phenotypic drug screening involves testing compounds on an organism level to identify potential leads

Example: Drug screening on an antibiotic-resistant bacterial strain to identify potential new leads

LBDD uses known compounds to guide drug discovery

Ligand-based drug design (LBDD) relies on the properties of known bioactive compounds

Motivation: If we find compounds with little bioactivity, we can use LBDD to find compounds with similar chemical features to improve specific outcomes

Assumption: Similar structures can lead to similar—hopefully improved—biological effects

LBDD does not **require** the structure of the target protein, making it useful when this is unknown

Key differences between structure- and ligand-based drug design

Structure-Based Drug Design:

- Requires 3D structure of the target protein.
- Uses the binding site structure to model potential interactions.
- Often employs docking and molecular simulations.

Ligand-Based Drug Design:

- Requires no structural information of the target.
- Uses the chemical structure and activity of known ligands as guides.
- Relies on molecular similarity rather than direct binding predictions.

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Chemical space exploration is still challenging, and now we need to identify similar compounds

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After today, you should have a better understanding of

How descriptors and fingerprints evaluate molecular similarity.

Quantifying molecular similarity is challenging

Suppose we performed an experimental high-throughput screen and identified these **potential leads**

Which group of molecules should we pursue for increased bioafinity?

With your neighbors, determine how you would choose the group of molecules to pursue.

Computed with SwissADME

Molecules can have similar properties, with slight structural differences causing widely different functions

Simple descriptor comparisons are not sufficient for computing molecular similarity

Computed with SwissADME

Molecular fingerprints encode structural information

Extended Connectivity Fingerprints (ECFPs) encode structural features into numerical representations

Phenylephrine

- 1 from rdkit import Chem
- 2 from rdkit.Chem import rdFingerprintGenerator
- 3 fmgen = rdFingerprintGenerator.GetMorganGenerator(
- 4 radius=3, fpSize=1024,
- 5 atomInvariantsGenerator=rdFingerprintGenerator.GetMorganFeatureAtomInvGen()
- 6 **)**
- 7 mol = Chem.MolFromSmiles("C1=CC(=C(C=C1CCN)0)0")
- 8 print(fmgen.GetFingerprint(mol))

How do we compute this?

Hash functions are used to encode chemical information

6—

10=

"Encoding" is a computational term for transforming information in a numerical format for computers

For each heavy atom (i.e., not H), hash atom-specific properties

$$ID_0 = \mathrm{hash}(Z_i, V_i, C_i, R_i, \ldots)$$

 $ID_0 egin{array}{ccc} Z & ext{Atomic number} \ ID_0 & V & ext{Valence} \ Valence \ Iteration 0 \ identifier & C & ext{Formal charge} \ R & ext{Ring membership} \end{array}$

Let's look at carbons 6 and 10

Because of the same element and connectivity, they have the same ID_0

•••

id6_iter0 = hash((6, 3, 0, 1))
print(id6_iter0) # 7468469475583712974

•••

id10_iter0 = hash((6, 3, 0, 1))
print(id10_iter0) # 7468469475583712974

For each additional iteration of *n*, incorporate the hashes of connected atoms that are *n* bonds away

Next, encode the atom IDs that are exactly one bond away

Format: (IterationNumber, AtomID, BondOrder1, AtomID1, BondOrder2, AtomID2, ...)

```
id6 iter1 = hash((
    1, 7468469475583712974, # ID for atom 6
    2, 901285887933171736, # ID for atom 5
    1, 901285887933171736 # ID for atom 7
))
print(id6 iter1) # -1070477880882296059
id10 iter1 = hash((
   1, 7468469475583712974, # ID for atom 10
    1, 901285887933171736, # ID for atom 5
    2, 7468469475583712974 # ID for atom 9
))
print(id10 iter1) # 9113858623660175530
```

Repeat for all atoms while hashing n - 1 IDs

Each iteration encodes local chemical information into each atom's ID

We can repeat the process for larger *n*, which captures more chemical information at a (small) computational cost

We keep track of atom IDs at each iteration to encode multiple "levels" of chemical information

Iteration 0

[-96873481, -5237400, -608624, -40896092, 13106358, 39304191, 13106358, 39304191, 39304191, 39304191, 18495798, 18495798]

Iteration 1

[-12887828, 34836456, -82428984, -76182021, 57441373, 18535308, 36698099, -16062189, -71082609, -16062189, -13803757, -35226747]

Iteration 2
[-30242937, -22342045, -3701095, -83323106, -81401022, -79585126,
259777, -18164777, -83853893, -9624634, -63890015, -86218719]

Iteration 3

[24482285, -67056973, -1049934, 58183281, 9686245, 65319696, -89546467, 90525418, -96278682, -31838946, -41820336, -42202112]

Iteration 0

[39304191, 39304191, 13106358, 13106358, 39304191, 13106358, -608624, -608624, -2248911, 18495798, 18495798]

Iteration 1

[-16062189, -16062189, -54942758, -54942758, 18535308, 80518135, -46276084, 85303560, -4225841, -13803757, -13803757]

Iteration 2

[45202524, -32527659, 91315393, -86313403, 74663225, 43056615, -92441264, 61456743, 35268850, -86729888, -86729888]

Iteration 3

[17051553, -83857497, -10864101, 42020134, 84228020, 88509243, 53634925, 58427327, 85169475, -62345869, -23012595]

Similar structural features will share atom IDs

until our iteration starts incorporating different structural features

Atom IDs are encoded into a bit array

We can get a collection of atom IDs, but how would we rapidly compare molecules with different number of atoms?

We use **bit arrays**, which are fixed-length collections of ones and zeros 10101100 11011010

This allows efficient operations	AND	10101100 11011010 10001000	Features that are in both molecules
	OR	10101100 11011010 11111110	Features that are in either molecules

Converting atom IDs to bit arrays

Decide on length of bit array, for example, 1024 and fill with zeros

Divide each atom ID by the length of the array and determine the remainder

Set the value of the bit array at that index to 1

dex to 1 ecfp[908] = 1

-1070477880882296059 8 1024 = 908

Tanimoto similarity compares the ECFPs between two molecules

Molecular similarity: The concept that similar molecules often show similar biological effects.

Using bit operations, we can compute similarity using Tanimoto

$$ext{Tanimoto similarity} = rac{c}{a+b-c}$$

- *a* is the number of bits set to 1 in vector **A**.
- *b* is the number of bits set to 1 in vector **B**.
- *c* is the number of bits set to 1 in both vectors **A** and **B** (the intersection).

This formula measures the ratio of the shared features to the total number of unique features between two molecules.

Tanimoto similarity ranges

How similar does ECFPs and Tanimoto say these molecules are?

After today, you should have a better understanding of

How QSAR models predict biological activity based on molecular structure.

QSAR models link chemical structure with biological activity

Purpose: To predict the biological activity of molecules based on their structure.

Motivation:

- Reduces the need for experimental screening.
- Helps identify potential drugs quickly and cost-effectively.

Example: Predicting if a compound is likely to be an inhibitor of a target enzyme based on known inhibitors.

Types of QSAR Models:

- 1. Linear Models: Simple, interpretable, e.g., linear regression.
- 2. Nonlinear Models: Capture complex relationships, e.g., neural networks.

Developing a QSAR model follows systematic steps

- **Data Collection:** Gather biological activity and molecular data.
- **Descriptor Calculation:** Calculate numerical descriptors for each molecule.
- **Model Selection and Training:** Use machine learning to correlate descriptors with activity.
- **Model Validation:** Test model accuracy with independent datasets.
- Interpretation and Application: Use the model for predicting new molecules.

Linear regression models are simple but effective for QSAR analysis

Fits a linear relationship between descriptors and output

 $Y=eta_0+eta_1X_1+eta_2X_2+\dots$

- Advantages: Easy to interpret.
- Limitations: Limited to linear relationships; struggles with complex datasets.

Nonlinear models capture complex relationships in QSAR data

Examples of Nonlinear Models:

- Neural Networks: Capture complex, nonlinear patterns in large datasets.
- Random Forests: Effective for high-dimensional data, robust against overfitting.

Example: Predicting toxicity, where relationships between descriptors and outcomes are often nonlinear.

After today, you should have a better understanding of

The role of pharmacophore modeling in identifying essential molecular features for activity.

Where QSAR quantifies activity, pharmacophore modeling identifies critical molecular features for activity

Pharmacophore modeling defines the essential features needed for biological activity

A pharmacophore is the 3D arrangement of molecular features required for biological activity

Building a pharmacophore model requires multiple active compounds

Step 1: Align active molecules

- Identify common structural features
- Determine spatial relationships
- Consider multiple conformations

Step 2: Define feature locations

- Mark shared pharmacophoric points
- Establish distance constraints
- Set tolerance spheres

Before the next class, you should

Lecture 18:

Exam 02 Review

Ligand-based drug design

- Finish A07
- Study for exam